MAIZE GENETICS COOPERATION

NEWS LETTER

12

March 6, 1938

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Department of Plant Breeding Cornell University Ithaca, N. Y. MAIZE GENETICS COÖPERATION DEPARTMENT OF PLANT BREEDING CORNELL UNIVERSITY ITHACA, NEW YORK

November 17, 1937

To Maize Geneticists :-

Contributions of material for the annual Maize Genetics Cooperation news letter is hereby requested. Any new linkage data, methods, hypotheses, suggestions, or anything else that you think may be of interest to other maize workers will be incorporated in this news letter. Since it is desirable to have the information presented in a somewhat uniform system, it is suggested that you refer to some of the previous Co-op News order to be included in this News Letter your write-up. In received by the Co-op not later than January 15, 1938.

Several years ago when a number of maize geneticists found that they were unable to get their linkage data published in some of the leading journals, they conceived the idea of combining their relatively small papers into one larger paper and publishing collectively. This suggestion was approved by the Editor of Genetics, and the Secretary of the Maize Genetics Cooperation signified his willingness to collect the individual papers for the publication. But to date only one paper has been received by the Co-op. Perhaps the reason for this lack of response from maize workers has been due to some misunderstanding of the plan. With this possibility in mind, it may be advisable to quote from a recent letter to Dr. Dunn, Editor of GENETICS:

"The Maize Genetics Cooperation circular letter does not constitute publication and none of the material in it may be quoted except by permission of the author. Much of the material in this Co-op News Letter is not complete, but rather is merely some ideas and indications which the men have obtained in their studies and are willing to pass on to other workers in this field to speed up progress with maize. Some of the material, however, is more complete and should be published so that it will be more readily available to other geneticists. This latter type of material will be included in the Co-op News Letter in the same form as in previous circular letters. But it will also be written up in a different manner to be included in the collective publication.

"The details of the method of handling the material in the proposed collective publication of linkage studies in maize will, of course, have to be worked out cooperatively between the publisher and the Maize Genetics Cooperation. During the several years that this idea of collective publication has been discussed among maize workers, the following plan has been formulated. Each cooperator who has linkage data which he considers useful and of permanent value to other geneticists, shall write a short paper in the same manner as he would if he were to publish independently. Then each of these papers will be sent to the Secretary of the Maize Genetics Cooperation, who will group them into one larger paper with an introduction, etc., and will serve as author of the collective paper. The important point is that each short paper will be an individual and separate unit within this larger paper, with the name and address of the author affixed to it. The Secretary of the Maize Genetics Cooperation shall be responsible for the organization and composition of the whole collective paper, but the respective authors of the 'unit papers' shall be responsible for their data. This means that any citation from the collective publication must include the name of the maize worker who furnished that particular data."

Dr. Dunn has written:

".....there is nothing in the policy of GENETICS to interfere with publication of maize linkage data in the form you suggest. Our numbers early in the year are likely to be the heaviest so May or July publication would fit our schedule best. Submission of the first paper in February would be most convenient for us."

It is suggested that you write your contribution to the News Letter first; then excerpt certain linkage data from it and write a separate papers(s) to be included in the collective publication. The particular data which you select for publication will appear in both the News Letter and the group publication. For further information concerning the general form of a linkage paper, see the Co-op News Letter of March 4, 1936, page 2; or March 23, 1937, page 15.

Sincerely yours,

D. D. Langham D. G. Langham,

Secretary

MAIZE GENETICS COÖPERATION DEPARTMENT OF PLANT BREEDING CORNELL UNIVERSITY ITHACA, NEW YORK

January 22, 1938

To Maize Geneticists :-

A number of maize geneticists have already sent in their items for the annual Co-op News Letter, and many of you probably have your contributions in the mail now. The <u>final date</u> for the receipt of material for this 1938 Letter is <u>January 31st</u>.

In the circular letter of November 17, 1937, I discussed the proposed collective publication of linkage data in such detail that the cardinal points were apparently lost in the shuffle. In brief, the plan is that linkage papers, any one of which in itself would not be sufficient for separate publication, will be sent to the Secretary of the Maize Genetics Cooperation who will group them in much the same manner as in BIOLOGICAL ABSTRACTS and send them to the Editor of GENETICS for publication. Each unit paper must be written as if it were to be published independently. No alterations or additions will be made by the Secretary of the Co-op.

In order to be included in this collective publication, your paper must be received by the Co-op not later than March 31, 1938.

Sincerely yours,

D. J. Langham D. G. Langham

Vol. 12

MAIZE GENETICS COÖPERATION DEPARTMENT OF PLANT BREEDING CORNELL UNIVERSITY ITHACA, NEW YORK

March 6, 1938

To Maize Geneticists:-

The material in this letter was obtained from many sources, and has been organized under the following heads:

- I. General News Items.
- II. Seed Stocks Grown in 1937.
- III. Seed Stocks Received For Propagation in 1938.
 - IV. Miscellaneous Co-op Items.
- V. Gene Index of all the Co-op letters.
- VI. Chromosome Maps of Maize.
 - A. Regular map: few genes, loci fairly definite.

B. Working map: many genes, loci not well established. Most of the information in this letter is given as it was received by the Co-op, but a few changes were made in some of the tables to conform to the accepted system of arrangement.

I. General News Items

University of Minnesota, St. Paul, Minn. -1. Zebra seedling, <u>zb_l</u>, has been located in chromosome 1 by the following studies.

Gene	8	Phase	XY	Xy	<u>xY</u>	xy	Total	% Recomb.
Zb4	Br	RS	448	142	152	12	754	31.1
Zb4	Fl	RS	455	135	158	9	757	28.0
Zb4	Bm2	RS	487	103	144	23	757	46.0
zb4	P	CS	266	24	5	64	359	6.9

Progeny of 1 ear indicated that the P parent was heterozygous giving the following segregation:

CS	63	30	2	24	199	6.7
		-				

2. A culture of rap received from Dr. Brink at Wisconsin proves to be similar to the one I have studied for many years. There is some variability in type of ear, some cultures showing rudimentary male flowers on the tips of some ears, irregularity of rows on the cob but no division of the cob as in ray. Other cultures have a divided cob on the tip of the ear but a solid cob at the base. Ra, can be separated from rap in the F2 of a cross.

3. Virescent seedling. A virescent seedling in Minn. #13 corn was found to be linked with japonica and given the symbol \underline{v}_{21} . Rhoades (Co-op News Letter, March 23, 1937) has found \underline{v}_{16} and \underline{v}_{21} to be allelic after trisomic tests had placed \underline{v}_{16} also in chromosome 8. Further linkage data of \underline{j}_1 , \underline{msg} and \underline{v}_{16} are as follows:

Genes	Phase	XY	Xy	xY	xy	Total	% Recomb.
J1 V16	RB	82	565	542	71	1260	12.1
J1 V16	RS	354	149	154	4	661	16.9
J ₁ Msg	CS	464	39	23	135	661	9.5
Msg V16	RS	337	150	171	3	661	13.9

The order of the genes appears to be $j_1 - ms_g - v_{16}$.

4. Zebra striped. Emerson et al list five cases of zebra striping that have been reported. There are two types, one that is expressed in the seedling stage and which may completely disappear in partly grown plants. The type reported here was obtained from an inbred line of Del Maiz sweet corn furnished by J. D. Barnard of the Minnesota Valley Canning Company. The season in 1936 was very hot and dry. Germination of sugary seeds was much lower than normal. Zebra striping could not be classified until late summer when the weather was cooler. Classification was difficult in some cultures. The results given in the summary indicate \underline{zb}_6 is located in group 4.

Genes	Phase	XY	Xy	XY	xy	Total	% Recomb.
Zb ₆ Tu	CS	410	64	64	90	628	23.3
Zb6 G13	RS	326	148	135	19	628	33.9
Tu Glz	RS	314	160	147	7	628	20.5
Zb6 Su1	CS	4227	259	175	361	5022	13.3

The order of the genes appears to be Su1 - zb6 - Tu - gl3.

H. K. Hayes and M. S. Chang

University of Missouri, Columbia, Missouri -

1. Of the unknown glossies grown in 1937, tests were completed on one which was found to be different from the other ten and has been assigned the symbol \underline{gl}_{11} . This was an X-ray induced mutant. One of the ultra-violet induced glossies proved upon test to be \underline{gl}_2 . Tests on three others have not been completed. In a previous report it was stated that somewhere along the line \underline{gl}_6 and \underline{gl}_g had been confused and the present stocks of these are identical. Since the symbol \underline{gl}_g has been used in print for the glossy on the 5th chromosome, this designation has been retained and a new glossy assigned the symbol \underline{gl}_6 .

2. Glossy 7 has been tested with $j_1 ms_g$ with no indication of linkage:

Genes	Phase	XY	Xy	XY	xy	Total	% Recomb.
G17 J1	CS	159	22	40	9	230	43
Gl7 Msg	CS	148	23	51	8	230	50

3. The inheritance of yellow endosperm color is more complex than has been generally believed. Evidence is available for the presence of at least one gene in addition to \underline{Y}_1 and \underline{Y}_3 which is concerned with the presence or absence of yellow endosperm pigment. Ratios of 3:1, 9:7, 15:1, 45:19 and others possibly more complex have been obtained.

The yellow scutellum gene sy is able to produce its effect in the presence of y_1y_1 , but in the presence of other recessive whites the development of pigment (carotin) is completely suppressed. The factor or factors involved have not been completely identified.

G. F. Sprague

John Innes Horticultural Institution, London, England -

1. Experiments on the inheritance of quantitative characters commenced by Dr. Brieger were continued during the summers of 1936 and 1937. The ultimate aim is to produce varieties of sweet corn which are early enough for the English climate and yet satisfactory in yield. In a comparison of F_1 families and their parents it was found that the application of a pseudo-factorial method of analysis (Yates, 1936) is not warranted for field trials with maize. The efficiency of the experiment when treated as a 3 x 3 x 3 pseudofactorial arrangement was about 60% of that when treated as a simple randomised block lay-out.

C. D. R. Dawson'

Connecticut Agricultural Experiment Station, New Haven, Conn. -

1. The evidence so far obtained indicates that mosaics in maize are due to losses or rearrangements of chromosome fragments rather than to somatic crossing over as Stern finds for Drosephila. Paired mosaics involving different chromosomes have been found for nearly all of the easily identified endosperm characters. In seeds heterozygous for \underline{C} and \underline{Pr} the following results have been obtained:

	White Spots	Red Spots	Red and	White	Paired	Spots
Number Ratio	8409 227	1061		37	,	

These figures indicate a more or less random exchange between the 60 chromosome arms in this triploid tissue.

The secondary paired mosaics (twin spots within the dark part of primary twin spots) can not be accounted for by somatic crossing over but are understandable on the basis of translocation followed by further breaking at other places. Variegated waxy tissue in areas that have previously lost the \underline{C} gene show an unstable condition that would not be expected from somatic crossing over. Similar variegation has been found involving \underline{C} , \underline{Pr} , and \underline{Su} .

In seeds resulting from the pollination of \underline{C} wx by \underline{c} Wx light and dark alcurone twin spots were found indicating a shift of one \underline{C} allele. If this resulted from an exchange of homologous segments the endosperm underlying the dark part of the twin spot should be waxy. In many such twin spots examined no waxy areas were found.

D. F. Jones

Agricultural Experiment Station, College Station, Texas -

1. The most important development in Texas during the past year is the discovery that the essential differences between Zea and Euchlaena are not due to numerous genes scattered at random over all the chromosomes as we first thought, but are due to four chromosome segments which are transmitted in inheritance in almost the same manner as single genes. The fact that these segments all carry genes similar to those possessed by Tripsacum, and the simultaneous discovery that short segments of the chromatin are interchanged between Zea and Tripsacum in hybrids of these two genera, has led us to the conclusion that teosinte is nothing more than maize with several translocation segments from Tripsacum superimposed upon the maize germplasm; the product of a natural hybrid between Zea and Tripsacum.

Two of these translocation segments have been located by linkage studies. They occur at opposite ends of chromosome 4 and both show linkage with <u>Su</u> and <u>Tu</u>. These translocation segments from Tripsacum are probably the cause of the unpaired terminal segments which Longley has observed in his cytological studies of the hybrid of maize and teosinte. We have verified his observations on the occurrence of these segments but we are not yet certain that they occur in every case on the chromosomes which he has designated.

The differences between the various kinds of teosinte which have been collected in Guatemala and Mexico may be attributed partly to the differences in the maize to which these translocations segments have been added, and partly to a loss of portions of one or more segments as the result of repeated hybridization with maize.

These new facts reopen the entire question of the origin of

maize. With teosinte as a recent development out of the picture, it is reasonable to assume that maize originated from pod corn, which in the homozygous condition is frequently a perfect flowered plant similar to the Andropogonae, and which has the essential characteristics of a plant adapted to survival in the wild. The place of origin was probably in South America, either in Peru or Bolivia.

We suspect that the crossing of South American types of maize with Tripsacum to produce the new genus Euchlaena, has also resulted in some new types of maize previously not in existence, such as the pointed pop corns and the long slender flint and flour corns, neither of which are known in Peru or Bolivia. If this is the case most of our North American maize varieties, with the possible exception of the Southern Gourd-seed types, carry Tripsacum genes in their germplasm. It is possible that the knobs which many of our North American corn exhibit on the chromosomes have been received from Tripsacum via Euchlaena, in which case we are quite likely to find some South American varieties which are lacking in knobs.

These hypotheses suggest a number of genetic and cytological tests which will keep us well occupied for a number of years. We are having some difficulty in locating viable seed of Bolivian and Peruvian maize and if any of the readers of this letter have such seed available we should appreciate receiving some of it. P. C. Mangelsdorf and R. G. Reeves

Iowa State College, Ames, Iowa -

1. The following linkage data were obtained from the backcross:

 $\frac{+ + Kn +}{br f_1 + bm_2} x br f_1 kn bm_2$ $\underbrace{0 \quad 1 \quad 2 \quad 3 \quad 1,2 \quad 1,3 \quad 2,3 \quad 1,2,3}_{165 \quad 182 \quad 14 \quad 4 \quad 50 \quad 50 \quad 47 \quad 52 \quad 7 \quad 10 \quad 1 \quad 2 \quad 16 \quad 32 \quad 6 \quad 2 \\ 347 \quad 18 \quad 100 \quad 99 \quad 17 \quad 3 \quad 48 \quad 8 \\ 2.8\% \quad 15.6\% \quad 15.5\% \quad 2.7\% \quad 0.5\% \quad 7.5\% \quad 1.3\%$

Recombination percentages: br-f1 7.2, br-Kn 26.1, br-bm2 35.2, f1-Kn 27.0, Kn-bm2 24.1.

These data do not agree completely with the present accepted location of <u>br</u> and <u>fl</u>. On the basis of these data <u>Kn</u> is located closer to <u>br</u> than to <u>fl</u> but it must be between <u>fl</u> and <u>bm</u>₂. After more extensive tests in 1937 the writer is doubtful that homozygous knotted plants can be distinguished from the heterozygous plants.

2. A tall late type of plant with about 50 per cent more nodes than the normal was discovered among the plants from an F_2 selfed

ear from the Krug variety. Plants of this type were crossed with several normal stocks in 1936 and the F_1 progenies were grown in 1937. All of the F_1 plants were normal. A similar type was found in 1936 among the plants from another F_2 selfed ear from the Krug variety.

A. A. Bryan

California Institute of Technology, Pasadena, Calif. -1. Correlation between cytology and map position in chrom. 1.

Cytological Position	Linkage Map Position
S .7	near sr
S .6	near P
S .4	$P \pm 1.5$
S .3	P = 7.6 - T - DT
S .25	P - 1(.3 - T - br)
	P - 20.0 - T - 34 - br
	P = 25.6 - T - 32.4 - br
	P - 24.2 - T - 27.8 - br
L .25	P - 39.0 - T - 3.8 - br
L .2	br - 13.4 - T
	near br
L .3	near br (± 2.7)
L .4	near br
L.4	near br
L .6	near br
L .6	br + 7.0
	near ad and an
	$br - 9.7 - T - 39 bm_2$
	br - 20 T - 45 bm2
L.S	br - 34.7 - T - 18 bm2
	Cytological <u>Position</u> S .7 S .6 S .4 S .3 S .25 L .25 L .2 L .3 L .4 L .4 L .4 L .6 L .6 L .8

2. Chocolate. In the distal part of long arm of chrom. 2. Homozygous long inversion gave the linkage order: $lg_1-44-v_1-32-B-25-Ch$

As the inversion includes about 4/5th of the long arm, <u>Ch</u> must be very near the end.

E. G. Anderson

3. Ms₂₀. Backcross tests with the following chromosome alterations show no obvious linkage:

Inversion of chrom. 2 (near <u>B</u> and beyond <u>v</u>₄) T2-4b (2 near <u>v</u>₄, 4 beyond <u>gl</u>₃) T2-3c (2 near <u>sk</u>, 3 near <u>d</u>₁) T4-8a (4 near <u>su</u>, 8 near spindle attachment)

4. Correlation between cytology and map position in chromo-

some 2.	Cyto _Por	ological sition	Linkage Ma Position	rb	
T2-3a T2-6b T2-9a T1-2b T2-3c T2-3c T2-3d T2-4d T2-9b T2-7b T2-7b T2-7b T2-7c T2-7c T2-7a T2-7a T2-7a T2-7a T2-7b T2-4b T2-4c	SSS SLLLLLL	•75 •6(?) •1 •1 •25 •2 •3 •3 •3 •3 •6+	near 1g B + 2.2 B - 2.7 B - 5.3 B - 13.0 B - 15.0 B - 15.	T - 23.7 - T - 30.9 - T - 28.0 - T - 12.4 - T - 12.4 - T - 7.0 - T - 7.0 - T - 7.0 - T - 7.0 - T - 4.6 - T - 7.1 - T - 6.0 -	v4 v4 v4 v4 v4 v4 v4 v4 v4 v4 v4 v4 v4 v

I. M. Clokey and E. G. Anderson

5. Linkage of <u>sb</u>. Slit blade is probably not on chromosome 6 where first reported:

Genes	Phase	XY	Xy	<u>Yx</u>	xy	Total	% Recomb.
Y ₇ Sb	RS	930	249	306	84	1569	50
Pl Sb	RS	478	135	154	49	816	48
Sug Sb	RS	896	265	340	68	1569	43
Py Sb	RS	1165	328	352	68	1913	50

The Y was not certainly, but probably, \underline{Y}_1 . In any case, \underline{sb} is not between \underline{Y}_1 and \underline{py}_1 .

6.	<u>Sb</u> is not	on chron	nosome a	2.			1
Genes	Phase	XY	Xy	<u>Yx</u>	xy	Total	% Recomb.
Lg1 Sb	RS	288	95	76	35	494	50
Gl, Sb	RS	288	92	78	36	494	50
sb x tr	isome 2:						

		Sb	SD
Culture	1	6	3
Gulture	22	10/	20
Curture	2	319	86

If sb were on chromosome 2 there should be about 30 sb plants; if on some other chromosome, about 100.

Notes: Sb is generally readily classifiable, though quite variable. Many of the plants are fully fertile. Usually the ratio is about as expected, though in two of my cultures the ratio was about 3:1 (F2 seed). This was not due to lethalness of sb, for nearly all of the seeds grew. In A B Pl plants the slitting of the blades seemed less developed than in green plants.

J. Shafer

University of Wisconsin, Madison, Wisconsin -

1.	Linkage of	ra2.					
Genes	Phase	XY	Xy	XY	xy	Total	% Recomb.
Cr. Rao	RB	14	26	22	3	65	26.1

This is further evidence indicating that the rap locus may be near that of dy.

R. A. Brink

Arlington Experiment Farm, Arlington, Virginia -1. The dominant Dt gene has been reported (1936) to produce dots of aleurone color on a1-tester seeds. The nature of the interaction between Dt and a was unknown at that time. It has now been established that Dt causes al to become unstable and to mutate at a rate thousands of times greater than normal. Muta-tions of al in the presence of Dt can be detected in aleurone, husks, and leaves i.e. plant color, and pericarp tissue. Recessive \underline{a}_1 mutates to the \underline{A}_1 allele a thousand times as frequently as to the ap allele. There is no chromosome abnormality present in the Dt line. The al gene is in chromosome 3 while Dt may belong to chromosome 9. Mutations of \underline{a}_1 to \underline{A}_1 or \underline{a}_1^p occur late in development in all tissues. It is not possible, at least by the writer, to reconcile these data with any of the hypotheses advanced by Schultz, Stern or Patterson to explain varicgation. They seem, however, to agree with Demerec's conception of increased mutability being caused by a chemical or physiological condition produced in the cell. Recessive al is highly stable in the presence of dt. The Dt gene is specific in its effect on a1. No other recessive locus including \underline{a}_2 , \underline{c} , \underline{r} , \underline{lg}_1 , \underline{wx} and \underline{su} is affected. A dominant modifying gene reducing the 'frequency or rate of mutation has been isolated. There is some evidence of a recessive gene affecting the time of mutation.

2. The following data on the location of wsz show the order to be as follows:



These four genes are all located in the short arm of chromosome 2 and if the RS or rS alleles are used with B all of them can be classified in the seedling stage.

	ws ₃ lg ₁ + - 415	ws3	lgl	gl ₂ -	16
selfed	+ + gl ₂ - 525	+	+	+	1068
	wsz + gl ₂ - 24	ws3	+	+ -	90
	+ lg ₁ + - 99	+	lg1	gl ₂ -	4

Total = 2241

 $ws_3 - 1g_1 = 11\%$ $ws_3 - gl_2 = 27\%$ $lg_1 - gl_2 = 19\%$

 $\frac{ws_3 lg_1 +}{+ gl_2}$

3. Trisomic tests show \underline{v}_{10} is in chromosome 6. Since \underline{v}_{10} gave 43% recombination with \underline{Y}_1 it will fall near the end of either the long or short arm. Tests with \underline{py} will be made this spring.

4. Preliminary results indicate that the pollen tube is not parasitic but is dependent for its growth in the silk upon the starch stored in the pollen grain.

5. There is a highly significant increase in crossing over in the $\underline{A_2}$ -Bt and $\underline{Bm_1}$ -Pr regions of chromosome 5 in microsporocytes as compared with megasporocytes. In a "low" line there was 7.6% recombination between $\underline{A_2}$ -Bt in the female gametes contrasted with 12.2% in the male gametes. Similar differences between the frequency of crossing over in the two sexes is the explanation of the inexplicable difference found by the writer (1936) in crossing over for the $\underline{Bm_1}$ -Pr region in plants hyperploid for the short arm of chromosome 5 as compared with diploid sibs. The hyperploid individuals had been used as the male parent while the diploid sibs had been used as the female.

M. M. Rhoades

Cornell University, Ithaca, New York -

1. In the News Letter of March 23, 1937, pp. 1, 2, it was shown by means of three-point tests involving the genes \underline{sr} , \underline{P} , and \underline{br} and the translocations TL-5a and TL-5c, that the order of the genes is $\underline{sr} - \underline{P} - \underline{br}$ with the translocation breaks between \underline{P} and \underline{br} . Backcross data from 476 individuals were also presented suggesting that \underline{ts}_2 is between \underline{P} and what was then called TL-10b but now designated TL-2c.

Records of the past summer presented below show that TL-2c is to the left of <u>P</u> very near <u>sr</u>, that <u>ts</u> is to the left of <u>P</u> with <u>ms</u>₁₇ presumably to the left of <u>ts</u>₂, and that TL-3a and TL-9c are probably to the right of P. The data are as follows:

Genes	Phase	XY	Xy	<u>xY</u>	xy	Total	% Recomb.
Sr T1-2c	CB	151	1	1	144	297	0.7

F ₁ genotype	0	1	2	1,2	Total
$\frac{ts_2 P +}{t+1} + T1 - 5b$	76 93 169	1 2 3 1.4%	17 22 39 13.7%	0 0	211
$\frac{ms_{17}P}{++T1-5b}$	26 25 51	3 2 5.7%	15 13 28 32.2%	2 1 3.4%	87
$\frac{ts_2}{t} \frac{P}{t} + \frac{T}{t} + \frac{T}{3}a$	106 140 246	3 1 4 1.3%	29 36 65 20.6%	0 0	315
$\frac{ms_{17} P +}{+ + Tl - 3a}$	54 33 87	2 0 2.0%	7 5 12 11.9%	0 0	101
$\frac{sr}{+} \frac{P}{+} \frac{+}{+} T1-9c$ From ¹ 37 News Letter	³⁸ 32 70 <u>97</u> 167	24 17 41 35.0% <u>24</u> 65 26.4%	0 5 4.3% <u>5</u> 10 4.1%	0 1 0.9% 3 1 1.6%	117 129 246
$\frac{T1-2c + P}{t ts_2 +}$ $\frac{ts_2 P}{T1-2c + +}$ From '37 News Letter	156 138 294 204 276 <u>480</u> 774 <u>401</u> 1175	51 29 50 49 <u>99</u> 179 18.7% <u>70</u> 249 17.4%	2 1 3 0 3 0 .6% 11 0.8%		377 583 960 476 1436
$\frac{+ ms_{17} P}{T1-2c + +}$	152 157 309	29 31 60 15.7%	3 5 2.0%	0 4 4 1.0%	381
Two of the culture chromosome 2 and data from \underline{B} $\underline{T1-3}$ + +	res reporte $\frac{P}{2c}$ and $\frac{ts_2}{P}$ $\frac{ts_2}{ts_2}$ +	ed above in of chromos	nvolving Ti some 1, gav	L-2c with <u>B</u> re the follo	of owing
<u>0</u> <u>1</u> 122 111 27 34	2	<u> </u>	<u>1,3</u>	<u>2,3</u> <u>1,2</u> ,	<u>3 Total</u>
233 61 16.2%	51 13.5%	2 29 0.5% 7.7%	1 0.3%		377

One of these cultures also segregated $\underline{lg_1}$ as in F₂. Using only $\underline{lg_1}$ plants, the records for $\underline{lg_1} = \underline{T1-2c} + \underline{P}$ are: $+ + + \underline{ts_2} + \underline{rs_2}$

0	1	2	3		1,2	1,3	2,3	1,2,3	Total
28	15	12	10	0	67.6%	3.8%	1.3%	4 5.1%	79

One of the cultures reported above to show close linkage between T1-2c and \underline{sr} also involved \underline{B} of chromosome 2 but no marker other than \underline{sr} of chromosome 1. The data are:

F ₁ genotype	<u>0</u>	1	2	1,2	Total
$\frac{B T1 \rightarrow 2c +}{+ + sr}$	63 44 107	28 9 37 25•7%	0 0 0	0 0 0	144

Since no crossover between T1-2c and <u>sr</u> appeared in this culture, the orientation of these two markers with respect to the rest of chromosome 1 cannot be told.

2. An	nong 2052	F2 plant	ts of	crosse	s of ad	y with an	, no double
recessive	appeared,	but F3	cultu	res fr	om 220	F2 an1 an	d ad1 plants
indicated	a crossov	er value	e of 4	.1% (L	inkage	Summary,	1935, p. 32).
Backcross	cultures	of last	summe	r gave	the fo	llowing r	esults:
Genes	Phase	XY	Xy	XY	xy	Total	% Recomb.
Adl Anl	CB	247	7	10	199	463	
Ad, An,	RB	4	36	31	1	72	

R. A. Emerson

4.1

 3. Chromosome 7.

 F_1 genotype
 0
 1
 2
 1,2
 Total

 in + + 880
 30
 99
 8
 1017

 in + + 980
 30
 99
 8
 1017

 $in - 4.3 - v_5 - 12.2 - gl_1$ $in - 4.3 - v_5 - 12.2 - gl_1$ 123

 $in gl_1 +$ 81
 12
 23
 7
 123

 $in - 15.5 - gl_1 - 24.4 - ij$ $in - 15.5 - gl_1 - 24.4 - ij$ $in - 15.5 - gl_1 - 24.4 - ij$

A number of seedlings in the latter cross were destroyed by mice in early stages. Counts are not dependable for distances but they are consistent with the order in the first cross.

A. C. Fraser

4. Doubling the number of chromosomes in yellow corn increased the carotinoid content 43 per cent as determined by chemical analysis of 2N and 4N stocks having a common origin. The volume of the endosperm cells of the tetraploid was more than 3.5 times as great as that of the diploid. Thus the individual endosperm cells of the tetraploid contained more than 5 times as much carotinoid as did those of the diploid and in terms of gene concentration within the endosperm tissue the amount of carotinoid was increased 2.5 times as a result of chromosome doubling. Chemical analyses by D. B. Hand.

5. The following results have been obtained to date on haploid frequencies in seedling progenies from untreated and x-rayed (1500 r-units) pollen:

	<u>N</u>	<u>2N</u>	N/1000
From untreated pollen	66	126,308	.53/1000
From x-rayed pollen	31	24,619	1.25/1000

The haploids were identified with the aid of recessive seedling genes, stomate examination and root-tip chromosome counts. L. F. Randolph

6. The following characters have appeared in inbred lines: co - Corrugated leaf. Raised striations of tissue in seedling and mature leaves. Classification good. Viability normal. bky-Brittle stalk. Similar to brittle stalk-1. Classifica-

tion good. Viability normal.

good. Viability good in germinator, but hasn't been tested under field conditions.

fy-Fine stripe. Plant striped in seedling stage and throughout development. Classification good. Viability normal.

Pu_x-Purple plumule. Similar to Pu₁. w_x -White seedling. Similar to w_1 . w_x -White sheath. Similar to w_3 .

R. G. Wiggans

7. White seedling-1 (w1) has been known to be loosely linked with the Y1 gene of the sixth chromosome (Linkage Summary, 1935). To place wi more accurately in the chromosome seedling counts were made of the F2 cross between w1 and pigmy (py). Seeds were taken from the Co-op stocks. The results indicate a very close linkage between py and W1.

Genes	Phase	XY	Xy	<u>Yx</u>	xy	Total	% Recomb.
Py W1	RS	867	416	443	0	1726	4.8(if one xy)
					G. 4	A. Lebede	ff

8. Segregation in autotetraploid maize. To determine the nature of segregation of some genes in autotetraploid maize, backcrosses were made involving the genes <u>B</u>(plant color booster) and Su(sugary endosperm).

Cross	В	b	No. of	Ratio
and a second second second second			Plants	
BBbb x bbbb	437	135	572	3.25:1

Some difficulty was encountered in classifying the progeny of the backcross, sun red (BBbb) x green (bbbb), since there was a great deal of variation in degree of coloration. Some plants were distinctly sun red, others resembled dilute sun red, while still others showed a tinge of color on and around the ligules. Undoubtedly errors were made in classification, there being an excess of green plants. However, the backcross ratio approaches 3.67 : 1. Since the type of segregation is a function of cross over distance between the gene locus and the spindle fiber attachment region, this would indicate that the gene <u>B</u> is located fifty or more units from the spindle fiber attachment region and that chromatid segregation had occurred.

Cross	Su	su	No. of	Ratio
			plants	
Su Su su su x su su su su	2877	645	3522	4.46 : 1
su su su su x Su Su su su	<u>369</u> 3246	<u>87</u> 732	<u>456</u> 3978	4.24 : 1 4.43 : 1

There was no difficulty in classifying sugary segregates in a backcross of autotetraploids. The ratio of 4.43 Su : 1 su indicates that this gene has segregated on a basis intermediate between the random distribution of four chromosomes and random distribution of eight chromatids, and suggests that the gene Su is located about 20 cross over units from the spindle fiber attachment region.

H. E. Fischer

9. It has been observed by many investigators that the F_1 ears of maize-teosinte hybrids are 4-rowed (paired spikelets, two-ranked). This indicates that the paired spikelet condition

of the maize ear is dominant to the single spikelets of teosinte. Collins and Kempton, 1920, showed that in an F_2 population, paired and single spikelets segregated 3:1. Data obtained by the writer in the summer of 1937 have confirmed their findings.

It has not been pointed out, however, that the two-ranked condition of teosinte, which appears in the F_1 of maize-teosinte hybrids, segregates as a unit character in the F_2 population. The combined 3:1 segregation of the dominant two-ranked condition of teosinte (as contrasted with the many-ranked condition of maize) and the 3:1 segregation of paired vs single spikelets, gave a 9:3:3:1 ratio, indicating that these two genes are independent of each other. This independence makes possible the combination of the recessive many-ranked condition of maize with the recessive single spikelets of teosinte, giving two kinds of ears: some with an even number of rows and others with an odd number of rows. Thus, 3-, 4-, and 5-rowed ears with single spikelets have been found. With paired spikelets these would presumably have been 6-, 8-, and 10-rowed ears, respectively.

10. Preliminary F_2 and reciprocal backcross data on maizeteosinte hybrids indicate that response to short-day may be due to one, or a few, genetic factors.

11. New characters.

gc₂ - Glucostacious-2. Seedlings pale green in very early stages. Then brown blotches appear and the plants die. Chrom. unknown.

 cz_x - Cuzcoid. Plant too late to shed pollen under field conditions at Ithaca.

lat - Lazy teosinte. Similar to lal in maize. Has not been tested for allelism.

D. G. Langham

II. Seed Stocks Grown, 1937

1. Testers. Chromosome 1:

p ad1 seg. an1

 $P/ + f_1 bm_2 seg. br$

(p ad₁ x p ad₁ an₁)self sr an₁ bm₂

br f₁ bm₂ x (Kn x br f₁ bm₂)

Chromosome 2:

lg, gl, B ts, v4 A Pl x lg, +/gl2 B +/ts1 v4 A Pl

lg1 b gs2 V4/? Gl2/? x Inbred II

lg, Gl_/? b vh gs, x Inbred I

Chromosome 2 (con't): Inbred x lg, gl, b v4 A pl b gs2 lg1 lg1 g12 b v4 A1 Y trisome #2 Chromosome 3: a1 1g2 Dt/? al na cr gl v5 Y aj Dt/? a₁ Dt/? seg. 1g₂ a₁ +/na +/lg₂ +/ts₄ x a₁ na +/lg₂ +/ts₂ $A_1 \lg_2 + d_1 x A_1 \lg_2 ts_4 d_1$ al lg, d, x Al lg, d, ts4 a₁ lg₂ Dt/? y₁ seg. na al 1g2 ra2 na $ts_4 x$ na $+/ts_4$ al yt seg. na $a_1 ts_4 + /na Dt/? x a_1 + /ts_4 na Dt/?$ Trisome #3 Chromosome 4: (su₁ x d_H) x (Tu su₁ x d_H) (gl₃ x su₁ j₂)self su^{am} du (Ga x su₇)self (sul gl3 x wl)self (+/w4 +/su1)self (Ts5 su1 x la su1)self (Ts5 su1 x wl)self (Ts5 su1 x la) x la su1 Trisome #4 Chromosome 5: Homo. A1 C R a2 bt bv pr1 Homo. A1CR a2 bt bv pr1 seg.v2 v2 a2 A7 CRbpl Homo. A1 CRA2 bt bv pr1 Trisome #5 Chromosome 6: Pl sm x pb_x(Lebedeff) Pl sm +/py A b x Pl py A b Y₁ Pl sm seg. py Chromosome 7: ral gll ij x bdl Inbred x raj glj ij blx ra₁ gl₁ ij x gl₁ ij fr₁ +/fr₂ ral gll ij v5 gl1 Tp seg. ra1 tp (bd₁ x gl₁ ij) x gl₁ ij bd₁ ral gl1 v5 x Tp gl1 v5 Trisome #7

```
Chromosome 8:
    v16 msg j1 x (msg j1 x v16)
    msg x msg/+
Chromosome 9:
    Inbred I x g4 wx
    g4 wx x (g14 x yg2 c sh wx)
    auy aug sh
     wx da ar saj
    Trisome #9
Chromosome 10:
     r zb5 seg. nl1
     Og/+ Y PWr
     Inbred x OgOg
     r A<sub>1</sub> C y<sub>1</sub> seg. g1
2. Miscellaneous
     U. S. 204 (Inbred I)
     Inbred I x bm3
     A, CR PI BY, a2
     g2 A1 b Pl
     seg. V12
     V13
     vaz x vaz/+
     wa x wa/+
     ms5 x ms5/+
     ms_6 \times ms_6/+
     ms7 x ms7/+
     ms_9 \times ms_9/+
     ms10 x ms10/+
     ms12 x ms12/+
```

```
msg j<sub>l</sub> x msg/+ j<sub>l</sub>
Trisome #8
```

Inbred I x ar wx c sh wx bp ms₂ x ms₂/+ (gl₄ x yg₂ c sh wx)self

l_l seg. w_l Og Og seg. l<u>l</u> Trisome #10

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West Branch (Inbred II)

seg. hf

Kn A<sub>1</sub> +/b Pl x A<sub>1</sub> +/b Pl

A<sub>1</sub> C R A<sub>2</sub> Pr<sub>1</sub>

(bm<sub>3</sub> x yg<sub>3</sub>)self

A<sub>1</sub> C R A<sub>2</sub> pr<sub>1</sub> i

Vg/+ x vg

an<sub>2</sub> x Inbred

+/na<sub>2</sub> x na<sub>2</sub>

r pr<sub>1</sub> x A<sub>1</sub> C R<sup>st</sup> B

A<sub>1</sub>B pl R<sup>st</sup> x r pr<sub>1</sub>

+/bk<sub>1</sub> x bk<sub>2</sub>

(+/bk<sub>1</sub>)self

+/de +/mi x de mi
```

+/an2 x +/an2 ms13 x ms13/+ Inbred x mi msyl x msyl/+ Wc Y7 ms37 x ms37/+ f_x(Wiggans) ms39 x ms39/+ dea 11 ms42 x ms42/+ deb 11 VI2 X VI2 Prl dec 11 seg. glio 11 $(sb x A_1 b pl +/y_1 su_2)sib$ CO 11 WSy y₁ su₂ seg sb Chlorophyll typespb Yellowish green Sx rather 1t. green sy medium to lt. green PCx dark green Ch/? seg. gl Ts3/+ v4/+ x Rg/+ nl2 Ts3/+ v1/+ x R g1 C sh wx gc2 Seed stocks from Australia grown by Shafer in Calif. for the Co-op: 3 different stocks of yellow-striped seedling. 5 different stocks of virescent seedling. crinkly. 3. Stocks too late to mature at Ithaca. From Krug: black pericarp brown pericarp seg. tassel seed branched ear bract in tassel seg. dwarf seg. defective endosperm oily spots rolled leaf seg. mealy variegated pericarp semi-dwarf striped leaves ragged ms x ms/+ seg. zebra seedling zebra leaves crinkly. From Mangelsdorf mottled dwarf seg. vpx

4. No germination. J33a X A c R sh wx B Pl sr an, bm2 +/V15 x +/V15 da auj aug sh ms15 x ms15/+ msu x msu/+ ms₂₇ x ms₂₇/+ gu ar say pky III. Seed Stocks Received For Propagation in 1938 J. Shafer, Ithaca, N. Y .:-1. VIQ T 1-2b x T 1-2b T 2-40 R. A. Brink, Madison, Wisconsin:-2. (pm x lg₂ d₁) sib (A₇ pm x a₁ lg₂) sib J. H. Kempton, Washington, D. C .:-3. fs 4. A. Tavcar, Zagreb, Jugoslavia:-Hs M. M. Rhoades, Arlington, Virginia:-5. (wsz lg] B A, pl x gl2) x (wsz lg] b A, pl x gl2) W. R. Singleton, New Haven, Connecticut:-6. zb_x f x ys rap? +/bay su₁ x +/10 gl3 v4 x lg1 gl2 b v4 rgACYSu V5? yellow x yellow ys_x (7 cultures) 7. R. G. Wiggans, Ithaca, N. Y .:-Pux dec fx

IV. Miscellaneous Co-op Items

1. Seed stock inventory. In March, 1937, an inventory of the genetic seed stocks in the Co-op collection showed that 148 of the genes reported in the Linkage Summary, 1935, were not in the seed trays here. A list of those 148 genes was included in the News Letter, March 23, '37, and several maize geneticists responded by sending in 16 genetic stocks.

In January, 1938, personal requests were sent to each of the 25 geneticists who, collectively, had first reported the remaining 132 stocks. We have learned that about 75% of those genes have been lost due to inviability of seed stocks.

2. Assignment of linkage groups. One of the topics discussed at a special meeting of maize geneticists at the A A A S meetings in Indianapolis, was the problem of linking workable genes and developing more desirable tester stocks. This is an important question because there are more than 50 suitable genes that haven't been linked and some of the chromosomes are poorly marked.

The plan previously outlined for linking new genes has not been fundamentally changed, but it may well be reviewed here. Each of the ten linkage groups in maize has been assigned to one, or more, cooperator who is charged with testing unplaced characters with his particular chromosome and building up suitable tester stocks. The following assignments have been made:

Chromosome	1.	Emerson.
Chromosome	2.	Rhoades and Clokey.
Chromosome	3.	Brink and Woodworth.
Chromosome	4.	Singleton and Brunson.
Chromosome	5.	Burnham.
Chromosome	6.	Stadler and Lebedeff.
Chromosome	7.	Jenkins and Fraser.
Chromosome	8.	Sprague and Perry.
Chromosome	9.	Eyster and Shafer.
Chromosome	10.	Lindstrom, Wentz, and Bryan.

When a new gene is found, a few seeds involving it should be sent to the secretary of the Maize Genetics Cooperation who will grow them in an increase block and obtain a liberal supply of seed for the central repository. Then the secretary will send a few seeds to each of the above geneticists who will test for linkage in his particular chromosome.

This system has been devised not to limit the number of workers who are trying to link new genes, but rather to insure the linkage of every workable gene.

3. More vigorous genetic stocks. During the summers of 1935 and 1936, a number of maize geneticists tested a group of inbred strains for disease resistance and general desirability. The two inbreds, U.S. #204 and West Branch Sweepstakes, seemed best suited to Ithaca conditions and have been selected for use in the Co-op. They have been designated as Inbred I and Inbred II, respectively, and are being used in crosses with weak genetic stocks to increase vigor and, by repeated backcrossing of the segregates to the inbreds, to obtain a more nearly homozygous chromosome complement. Later, the segregates from each inbred line may be crossed to get hybrid vigor.

Last summer 17 genetic stocks were crossed with both Inbred I and Inbred II.

4. Linkage maps. The linkage maps attached to this Letter were prepared from the data in the Linkage Summary and the data which appeared in the Co-op News Letters since the Linkage Summary was published.

> Sincerely yours, D. M. Longham D. G. Langham Secretary

V. Gene Index of Co-op News Letters

This gene index of the Co-op News Letters was made so that the information in the Letters which might be of value in connection with linkage studies could be more readily found. It includes mainly those genes about which some statement of linkage has been made in the Letters, and does not include those that are merely mentioned without any supplementary information. John Shafer.

ag (=ij): a1: 1-25-34, p. 5 (January 25, 1934, page 5) 11-24-34, pp. 13, 18, 11 1-23-33, pp. 3, 6 3-6-35, p. 3 12-18-33, p. 6 9-13-34, p. 8 1-23-33, p. 6 al: 3-4-36, pp. 7, 10, 11 3-23-37, pp. 8, 13, 14 3-6-38, pp. 8, 15 12-18-33, pp. 3, 5 1-23-33, pp. 3, 6 3-6-35, pp. 3, 5 3-4-36, pp. 15, 16 3-23-37, pp. 8, 15 a2: 12-18-33, p. 5 1-25-34, p. 6 11-24-34, pp. 2, 14 an: 1-23-33, p. 6 1-25-34, p. 4 11-24-34, p. 5 3-4-36, pp. 7, 17 3-6-38, pp. 9, 15 1-23-33, p. 6 3-6-35, p. 1 3-36-38, pp. 6, 11, 14 az: 11-24-34, p. 10 ara: 9-13-34, p. 2 ady: 1-23-33, p. 6 1-25-34, p. 4 ar: 3-6-35, pp. 3, 15 3-4-36, p. 9 12-18-33, p. 2 1-25-34, p. 8 1-23-33, p. 3 3-6-38, pp. 6, 11, 14 3-6-38, p. 16 ad2 (= ad7): as: 3-6-35, p. 3 1-23-33, p. 6 ad, (first called ad,): anj: 3-6-35, pp. 3, 15 12-18-33, p. 2 1-25-34, p. 8 1-23-33, p. 3 adz (now ad)) 3-6-35, p. 3 3-6-38, p. 16

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2 3 4 5 1 8 9 6 10 0 1003 0 | a, An 0 a2 6 bm, o de, 0/10 0 516 2 ×mot ofin ORP NS 11 4 lg, 12 br 14 mag 13 Y, 18 na Csh 21 24 gl, 22 28 P 28 may 24 71 gl2 pbx 30 28 li 28 31 pri Tp 32 AL Ja, 35 39 ba, ys, 38 ij to 40 39 Pl 41 43 91 49 B 53 as 51 sm 56 tay 56 sh 54 ww Te 5 56 51 R 56 Bn, 63 Rg 61 py bd, 68 te, 69 67 74 sp lou Bh fr, 66 14 74 ta, 72 12 gm4 frz deile ar 80 85 or may an ft Hs 82 VA f 85 di cr2 su, cb 02 al db dz 84 dy su2 893 baz gaz2 N6 day az sl Ch Daz2 va 102 an, gl 8 100 Tu NT def ds 103 cr, Wh de 15 105 82 oy wi du 92 Ype dz 1 re, WS 94 13 111 | gl3 Y re2 lg2 gm2 w6 gme la, sc, I l, yd may sf S, L6 129 90, l2 192 tr st ly Ng N3 l4 pm mez raz NP3 N12 lg mezo rt W4 NP2 nl, 03 yt we 156 Bm2 Y2 pp Og 356 Pr2 491 Pc2 ad, re3 pg, Kn moin sa sp2 tez sc2 N18 suz Tue N20 364 N15 up, will 392 W2 yf 32 xn, 305 Cinkage map of the ten chromosomes of zea mays showing the loci of those genes whose position can be determined

with reasonable certainty.

2 3 4 5 6 8 7 9 10 0 | por 0 a2 theme 5 10 12 o de, 0 1 000 0/100-2 1 Ar 0 Knob 2 492 0 0 1516 ORP 4 He 11 (23) al in 02 55 12 SAL 3 7 01 (me, 13 14 ma 47 the 2 des 16 16 Paz 17 16 Oq 21 main 25, ma 27--ta 28-5 P 20 22 az di glz 24 sh 21 24 71 27 si (ne li ma 25 28 na, de la 30 18 28 pt. 28 31 pr, gl, 22 35 364 28 Da, 35 Da. 39 36 39 43 ba, 28 28 1 39 6pe 1,5 40 42 41 41 PL 43 wl 32 36 Tp 902 45 47 lg2 va, 49 В 47 dz 38 ife 49 su2 sm 55, st 50 41 10,1 tay as 53 56-Ta 5 56 54 sk wi 56 59 KR 59 L 48 fr, 61 py 63 65 Rg 192 61 63 64 66 any 66 69 71 w fl, baz ta, sp. 20 68 12 74 ar 56 Bn NI 56 ±5 193 su, 72 12 Bh de 16 74 72 Pr2 + 20 75 152 et gm4 br 80 82 VA 36. gaz. 84 f, 156 85 85 d, 04 Nr2 84 dy Ny re, da, ch 4 AC de 15 WE ad, Tu vy 98 sf 100 91 Ad, who an, 102 84 100 l2 105 Y2 yd 103 02 12 Wh qme 491 Ly 111 la dz 111 93 may 2 de f ma 3 5, 03 raz f3 NP3 1392 129 wig rt pk qm2 132 Kn yt re 3 V18 N20 sa, rn, sly 365 yf 156 bm2 Ta, Linkage map of the ten chromosomes of zea mays showing the approximate loci of many genes. (Working map. 3- point more needed to teste establish exact loci of genes).

2 3 4 5 1 6 9 8 10 wa3 0 | a, o) de, 0 a2 6 km, 0 100 ofin 2 Knob 2 492 0 0 1516 ORP sn 4 15 lg, 12 br 11 14 mag 13 Y. 18 201 Csh 21 24 22 gl, 24 71 pbx 28 28 li 28 may, P gl2 28 pri 31 30 32 Tp Da, 35 38 4 to 40 ys, 39 41 Pl 39 ba, 43 91 49 B 51 sm wx 53 as 54 57 R 56 sh 56 tay 56 56 Bn, Tas 61 py Rg 63 bd. 69 67 74 spi loui de 16 Bh 68 fl, fr, 66 14 gm 4 72/12 frz 74 te, ar ft ma au, Hs 80 br 82 VA su, cb cr, 02 85 d, 85 f su2 dz 84 dy de al 893 N6 ba, gaz2 day az sl Da2 Ch gl8 va 100 Tu 17 des ds 102 an, Wh de 15 103 cr, w, 105 82 oy du Yp 92 re, 13 WS 94 dz 111 glz ne, Y gm2 gme w6 lg2 la, sc. l, I yd st may S, Lo l2 129 90, ton 192 st ly N3 Ng LA pm mez NP3 V12 lg raz mezo NP2 W4 rt nl, 03 yt wh Yz pp Og 156 Bm2 306 Pr, Pc2 491 ad, rez pg, Kn sa mon sp2 sc2 to2 N18 suz Tu N20 N15 304 vp. WII 392 W2 yf zl Xn, 305 inhage map of the ten chromosomes of zea mays showing the loci of those genes whose position can be determined

with reasonable certainty.